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ILLUSTRATED KEYS TO THE ANOPHELES MOSQUITOES OF THE PHILIPPINE ISLANDS

by

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FRANCISCO EDLAGAN BAISAS Dean of Philippine Culicidologists

То

we respectfully dedicate this work. His contribution to the knowledge of Philippine mosquitoes is without measure.

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INTRODUCTION

The first illustrated keys to the Anopheles mosquitoes of the Philippines were prepared by Russell and Baisas (1934, 1936). Periodically, since then keys have been produced (Simmons and Aitken, 1942; Bohart, 1945; Mendoza, 1954 a, b; Baisas and Bañez, 1957; and Baisas and Dowell, 1965).

The keys presented herein are an effort to produce simplified, well-illustrated guides to the identification of the Philippine anophelines, and have been tested extensively for the past two years on participant classes at the Malaria Eradication Training Center. The latest supra-specific concepts, especially those introduced by Reid (1968) are incorporated. Likewise, additional specific and sub-specific taxa are included.

Both the adult female and fourth instar larval stages have been dealt with in the keys. Captions for the key illustrations may be known by consulting the Table of Key Figures, page 1. The essential morphology of these forms, discussed in the succeeding sections, will enable the user to follow the keys successfully.

CLASSIFICATION OF PHILIPPINE ANOPHELES

The systematic index to the Philippine Anopheles species below follows the arrangement made by Reid and Knight (1961) and Reid (1968). It updates the checklists of Delfinado (1965) and Baisas and Dowell (loc. cit.). Unlike previous lists, here the former "groups" under subgenus *Cellia* are elevated to "series" which match the long-used series of the subgenus *Anopheles*. Furthermore, closely related species are arranged into "species groups", not "complexes" or "groups", as formerly done. With these changes the classification becomes much less complicated. Where trinomials are used in the paper, the third name indicates subspecies, i.e., *Anopheles gigas formosus* Ludlow. No scientific name lower than subspecies is recognized as valid.

One series name change is adopted here, i.e., *Pyretophorus* for *Pseudomyzomyia*, as suggested by Reid (1968).

The listing of the species groups to which the Philippine species belong, where applicable, serves to demonstrate the relationship of the local taxa to the Oriental anopheline fauna as a whole.

It was the opinion of Reid (1966, 1968) that A. subpictus Grassi may be absent from the Philippine Islands. Investigation of reared associated "indefinitus" from the saline, fish ponds near Manila has shown that some of these specimens are identical to the description of "subpictus" given by Reid. Likewise, A. vagus vagus Donitz has never been reported from the Philippines since King (1932) described the insular form, A. v. limosus. It has recently been collected from Mindanao Island by the authors. Therefore, these two are included in the keys and considered to be members of the Philippine fauna.

A new subspecies of *ludlowae* has recently been described by Darsie and Ramos (1969), and named *ludlowae cabrerai*. Reference was made to it by Baisas and Dowell (loc. cit.) and Reid (1968) as a variety of *ludlowae* which bears three dark spots on the anal vein instead of the usual two. It has been included in the following keys.

SYSTEMATIC INDEX

ANOPHELES OF THE PHILIPPINES

Genus Anopheles Meigen Subgenus Anopheles Meigen Anopheles series Edwards aitkenii species group acaci Baisas aitkenii James bengalensis Puri fragilis (Theobald) insulaeflorum (Swellengrebel and Swellengrebel de Graaf) lindesavi species group gigas formosus Ludlow lindesayi benguetensis King Mvzorhvnchus series Edwards albotaeniatus species group balerensis Mendoza eiercitoi Mendoza bancroftii species group ·pseudobarbirostris Ludlow barbirostris species group franciscoi Reid manalangi Mendoza vanus Walker hyrcanus species group lesteri Baisas and Hu peditaeniatus (Leicester) pseudosinensis Baisas umbrosus species group baezai Gater

samarensis Rozeboom Subgenus Cellia Theobald Mvzomvia series Christophers minimus species group filipinae Manalang mangyanus (Banks) minimus flavirostris (Ludlow) Neocellia series Christophers karwari (James) maculatus Theobald annularis species group annularis Van der Wulp philippinensis Ludlow Neomvzomvia series Christophers kochi Donitz kolambuganensis Baisas tessellatus Theobald leucosphyrus species group balabacensis baisasi Colless balabacensis balabacensis Baisas cristatus King and Baisas riparis riparis King and Baisas Pyretophorus series Edwards indefinitus (Ludlow) subpictus Grassi vagus limosus King vagus vagus Donitz ludlowae species group litoralis King ludlowae ludlowae (Theobald) hudlowae cabrerai Darsie and Ramos parangensis (Ludlow)

The Adult Female

MORPHOLOGY OF ADULT FEMALE

It is assumed that the user is already able to distinguish not only members of the Family Culicidae from other dipterous insects, but can also differentiate anopheline from culicine mosquitoes. If not, the reader is referred to Borror and Delong (1963) for the former and to Russell et al. (1963) or Delfinado (1966) for the latter.

The description below is by no means complete, but contains sufficient background to comprehend the key points. Certain structures need to be defined in order to understand the morphology.

 Sclerites — The integument of insects is made up of hardened plates called sclerites, separated either by lines, known as sutures, or membranes of various sizes. The body of the adult mosquito is composed primarily of sclerites, whereas the larval body is largely membranous. 2. Hairs and Scales – It is necessary to be able to differentiate between scales and hairs in adult mosquitoes. A hair is round, tapers from base toward apex, and it is movable, being connected to the body by a socket, called trichopore. The scales are flattened, immobile, without a trichopore, and usually widening, although the wing fringe scales are pointed, apically.

The females of Anopheles may be distinguished from those of other Philippine culicids by their long palpi, almost as long as the proboscis and by scutellum which is evenly rounded and beset by an unbroken, evenly spaced row of hairs posteriorly, see Fig. 1. Another characteristic is their living posture while at rest or taking a blood meal. Typically the body is held at a distinct angle from the resting surface and their thorax and abdomen form a straight line with the proboscis.

The Head

The spherical head, seen in Fig. 2, is about as wide as long, with compound eyes occupying a large portion of each side. Projecting forward are five appendages, the median slender proboscis, above which are the two palpi, and extending from the head between the eyes, the two antennae.

The proboscis is an elongate structure about onefourth the total length of the body. The visible portion is the labium, a sheath covering the piercing stylets. It is clothed with scales, usually uniformly dark with a light colored tip, the labella. In some species the apical half is wholly or partially beset with pale or flavescent scales.

The two, five-segmented **palpi** are also covered with scales, which are sometimes large, giving the structures a shaggy appearance. The scalation is either entirely dark or more often with pale banding. The terminal pale band is called apical; the next one, subapical, and the intervening dark band separating them is also called the **subapical dark** band. The number of pale bands may vary within one species, for the subapical dark band may be missing, or in others it may be found added where it normally is absent.

The antennae are composed of two basal elements, the small scape and bulbous torus. The remaining 14 rod-like segments, or flagellomeres, are collectively called the flagellum. Each flagellar part is adorned with a whorl of hairs, excepting the last, these hairs being much longer and more numerous in the male antennae. The Thorax

The thorax (Fig. 3) consists of three segments, called prothorax, mesothorax and metathorax. In mosquitoes, the mesothorax is greatly expanded at the expense of the other two. The dorsal aspect of the thorax is made up of



(Redrawn from Russell and Baisas, 1936)

the mesonotum (mn) and two smaller posterior sclerites, the scutellum (sc) and the postnotum (pn). The lateral portions of the anterior third of the mesonotum are somewhat depressed and are known as the fossae (fo). In most anophelines, scales are absent but hairs present on the dorsum of the thorax.

The lateral aspect of the thorax is known as the **pleuron**. It is composed of a number of sclerites which contain groups of hairs, or bristles. For the names and locations of the principal ones, see Fig. 3. The important sclerite and bristle group for the recognition of Philippine *Anopheles* females is the **propleuron** (prp) and its bristles (1).

Mosquitoes have one pair of functional wings attached to the mesothorax and one vestigial pair, called the halteres, on the metathorax. The former consists of a membrane supported by a network of veins. The veins have been named by the Comstock-Needham system in this key; see Fig. 4A for an illustration of the nomenclature. The wings are very important in the identification of Philippine anophelines. The ornamentation, especially the costal wing spots, is a salient feature. For an understanding of the pattern and terminology used here, see Fig. 4B.

Each of the six legs is composed of nine parts, as shown in Fig. 5. Leg characters are commonly used in the identification of Philippine anophelines. Various spotting and banding of pale and dark scales will be encountered. Spots may be confused with bands, so it is necessary to make sure the pale scales extend completely around the segment.

The Abdomen

The abdomen is composed of eight visible segments designated by Roman numerals I-VIII (Fig. 6), each consisting of a dorsal sclerite, the tergite (te), and a ventral one, the sternite (st). In most anophelinie females these abdominal sclerites have few or no scales; however, in some, scales or scale tufts may be present. These dorsal and ventral sclerites are separated by a membranous integument, the pleural membrane (pm). No pleural sclerites are present on the abdomen. The terminal segments constitute the female genitalia, and the lobe-like cerci (ce) are protruding at the posterior end.



Figure 2. Head of anopheline, showing details of morphology (60x).



Figure 3. Thorax of anopheline showing sclerites and bristle (hair) groups (60x). Legend:

cxl	coxa of prothorax
cxII	coxa of mesothorax
cxIII	coxa of metathorax
fo	fossa of mesonotum
ha	haltere
me	meron
mep	mesepimeron

- mn mesonotum pn postnotum prn pronotum pprn postpronotum prp propleuron sc scutellum
- st sternopleuron
- 1 propleural bristle
- 2 upper mesepimeral bristles
- 3 prealar bristles.
- 4 spiracular bristle





Figure 4. Wing of anopheline (44x).

À.	Venation by Comstock-Nee	dham	System:		
A C Cu h	anal vein costal vein cubital vein humeral cross vein	M m-cu R	medial vein medio-cubital cross-vein radial vein	Rs r-m Sc	radial sector vein radio-medial cross-vein subcostal vein
В.	Markings on hypothetical w	ing:			
A Ad as Bd bs f	apical pale spot apical dark spot accessory sector pale spot basal dark spot border scales fringe scales	fs H Hd P Pd Ph Phd	fringe spot humeral pale spot humeral dark spot preapical pale spot preapical dark spot prehumeral pale spot prehumeral dark spot	pi Psd S Sc Sd	pale interruption presector pale spot presector dark spot sector pale spot subcostal pale spot sector dark spot





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10(3).	Tarsomeres of hind legs with broad, pale bands, that on apex of third tarsomere extending over onto base of fourth (Fig. 30)	peditaeniatus
	Tarsomeres of hind legs with narrow, pale bands, fourth tarsomere without basal, pale band (Fig. 31)	
	+	
	r	
	Figure 30. (25x)	
	*	
х •	Figure 31, (26x)	
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	Coxae of mid legs and humeral cross-veins without scales (Figs. 34, 35)	lesteri
	Figure 32. (42x)	Figure 34. (50x)
	Figure 33. (46x)	Figure 35, (50x)









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19(18). Prominent tufts of dark scales present on sternites of abdominal segments II-VII (Fig. 58) . . . kochi Tufts of dark scales absent, or if present, only on sternite of abdominal segment VII (Fig. 59) . 20 Ţ T M Π ν ΔI VΠ Figure 58, (27x) Figure 59. (25x) 20(19). Tibio-tarsal joint of hind legs with conspicuous, broad, pale bands (Fig. 60) (leucosphyrus species group) . 21 Tibio-tarsal joint of hind legs with no pale bands, or at most narrow bands or spots (Fig. 61) . . . 24 Figure 60. (13x) Figure 61. (17x) 21(20). Presector dark spot of vein R with one or more pale interruption on at least one wing (Fig. 62); apical pale band of hind tibiae without longitudinal dark stripe (Fig. 63) 22 Presector dark spot of vein R without pale interruptions (Fig. 64); apical pale band of hind tibiae interrupted by longitudinal dark stripe ventrally or laterally (Fig. 65) 23 Figure 62. (28x) Figure 65. Figure 63. Figure 64. (38x)

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Figure 69. (22x)









Figure 93. (36x)

*Since A. sundaicus Rodenwaldt may occur in the Philippines, it would key to litoralis in this table. It may be distinguished from litoralis by the absence of the prehumeral pale spot on the wing.



*Second character does not apply to most of vagus vagus examined.

34(32). Subapical pale band of palpi one-third or less as long as subapical dark band (Fig. 99c) subapical pale band of palpi usually one-half or more as long as subapical dark band (Fig. 99d) subapical pale band of palpi usually one-half or more as long as subapical dark band (Fig. 99d)

Figure 99c. (30x)



Figure 99d. (35x)

The Fourth Instar Larva

MORPHOLOGY OF THE LARVA

The anopheline larvae are readily distinguished from other kinds of mosquito larvae by the absence of an elongate siphon on the eighth abdominal segment, and the presence of palmate hairs (Hair 1) on some or all of abdominal segments I-VII. The attitude of the live anopheline larvae when at rest on the surface of the water is characteristic, for they lie parallel to the top, whereas, most culcine larvae lie at a distinct angle to the surface.

The body of the mosquito larva is largely membranous, and beset by sclerites, such as the head capsule, abdominal tergal plates, spiracular apparatus and anal saddle. There are also numerous hairs attached both to the sclerites and to the membranes. The study of hairs (setae) is called chaetotaxy and the complete chaetotaxy of *Anopheles litoralis* King is depicted in Figs. 100-103. The nomenclature of Belkin (1951) is employed for the larval hairs. The morphology of individual hairs, shown in Fig. 104, is singularly important in identification of larvae.

The larva is divided into three distinct body regions; the head, somewhat flattened; the thorax, consisting of three fused segments, each indicated by its own set of hairs; and the **abdomen**, with eight obvious segments and two markedly modified structurally.

There are four larval instars, each separated by a moulting of the skin (ecdysis). A rapid, morphological identification of the four instars may be made by using the following simple key:

- a. Egg breaker present on frontoclypeus . . First Instar
- aa. Egg breaker absent b
 - b. Hair 16 of maxillary palpus

 - bb. Hair 16 of maxillary palpus present.....c
 - c. Imaginal eye absent Third Instar cc. Imaginal eye present Fourth Instar

The fourth instar is obviously the largest in size, but unless one is familiar with relative sizes of the species, it may be a poor character to follow. For instance, the third instar of *Anopheles manilangi* Mendoza may be larger than the fourth instar larva of *Anopheles filipinae* Manalang. The accompanying key is based on the fourth instar, but in most instances may also be used to identify the third instar.

The mature fourth instar larvae (pre-pupae) can be recognized by the appearance of the pupal hairs under their skin. The following detailed description applies to the fourth instar anopheline larva.

Head

The head is composed of a completely sclerotized capsule, Fig. 100. Three major plates are visible on the head, the frontoclypeus (fc), dorsally, and the two lateral plates (lp) which meet on the ventral side. A prominent line known as the epicranial suture (ep) separates the frontoclypeus from the lateral plates. Anteriorly are the antennae (an) and mouthparts. The antenna bears a hair on the shaft and several at its terminus. In some species the shaft hair (No. 1) is large, multibranched and located near the middle of the shaft. while in others it is small, simple and positioned in the basal one-third. The most conspicuous structures of the mouthparts are the mouth brushes (mb) terminal in position. The maxillary palpi (mxp) are prominent lobes just medio-ventral to the antennae, which bear subapical hairs (No. 16 of some authors: see Christophers, 1933 and Baisas and Dowell, 1965). On each lateral plate are found two eyes, a prominent imaginal eye (ie) and small, posterior, larval eye (le). The posterior border of the head is heavily sclerotized and called the collar (co). In early instars this collar is much wider.

Some of the 15 pairs of head hairs are of major importance in identification. The inner and outer clypeal hairs, head hairs 2 and 3, are widely used. The clypeal hairs may be simple, forked at tip, frayed, variously branched or dendritic, a term applied to branching which appears like the branches of a tree; see Fig. 104, A-D, L, M. The outer clypeal hairs are often difficult to see because they overlie the mouth brushes. It helps to view them with the 43x objective of a compound microscope. The same power will be necessary to see the frayings, or minute side branches, of the clypeal hairs. Less often used are the posterior clypeal and the sutural and transsutural hairs ---hairs 4, 8 and 9, respectively. Hair 16 on the maxillary palpi is not named by Belkin, but it is included for convenience since it is useful in larval instar recognition.

Thorax

The thorax is the thickest part of the body, and composed of three segments, the prothorax, mesothorax and metathorax. They are completely fused into one large body region, but the evidence of segmentation is found in the chaetotaxy, for there are three distinct set of hairs; see Fig. 103.



Figures 100-103. Fourth instar larva of *A. litoralis*, showing chaetotaxy. 100. Head, dorsal – left, ventral – right; 101. Palmate hair of abdominal segment IV; 102. Terminal segments of abdomen; 103. Thorax and abdominal segments I-VI, dorsal – left, ventral – right. *Legend*:

ag an atp co ct	 anai gili anterior tergal plate collar comb teeth 	ep – epicranial suture fc – frontoclypeus ie – imaginal eye le – larval eye lp – lateral plate mb – mouth brush	mxp ptp sa PRO MES MET	 – maxillary palpus – posterior tergal plat – anal saddle – prothorax O – mesothorax TA – metathorax
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Figure 104. Morphology of Hairs.

Certain thoracic hairs are used as diagnostic characters. The submedian prothoracic hairs 1, 2 and 3 show differences not only in the number of branches, but also the configuration of their bases. Sometimes their trichopores are set into sclerotized tubercles, which at times are joined together. Mesothoracic hair 4 (hair 5 of Puri, 1960, and Baisas and Dowell, loc. cit.) in one species has reclining, sinuous branches which form a star shape (stellate), whereas normally the branches are erect (Fig. 104. C, K). The pleural hair groups, hairs 9-12, on the ventral side of each thoracic segment are important because they differ in the number of branches among the various species. They are distinctive because each group of four hairs arises from a common tubercle. Metathoracic hair 3 may have hair-like or leaf-like branches. In the latter case it is known as the metathoracic palmate hair.

Abdomen

Following the interpretation of Snodgrass (1959) and Puri (loc. cit.), the abdomen consists of 10 segments of which the first seven are similar in composition; see Fig. 102. The eighth is modified to bear the spiracular apparatus, and the latter is composed of elements of both the eighth and ninth abdominal segments. Two sets of hairs, as interpreted by Belkin, on the eighth segment support the opinion that actually two segments are represented, VIII and IX. The terminal segment is the tenth, which ends in the anus, carries large dorsal and ventral tufts of hairs and possesses four **anal gills** (ag), thin-walled, tracheated lobes or **papillae**.

Each of the first eight abdominal segments has a sclerotized plate dorsally, called the anterior tergal plate (atp). Posterior to it on some segments are one or more smaller sclerites known as posterior tergal plates (ptp). The tergal plates may be difficult to see if the alimentary canal is filled with dense material. The tenth segment also has a large dorsal plate, called the saddle (sa). The larvae belonging to *Myzomyia* series have large anterior tergal plates, sometimes almost covering the entire dorsal surface of the segment.

The normal complement of hairs on the abdominal segments is 15 hairs, Nos. 0-14; however, VIII, IX and X have fewer; see Figs. 103, 102.

Certain abdominal hairs have decided importance as recognition characters. The palmate hairs (Hairs No. 1) are salient features on I-VII. Figure 104, E-J, show the various forms. The palmates may have leaf-like, flattened branches (E, G, I) or hair-like parts (J). The usual leaflet has a basal flat part, the blade, terminal, attenuated portion, the filament, and a section in between where a series of notches occur, usually with a more or less flattened area, known as the shoulder (G). Some species have the notches but no distinct shoulders. Others are without the attenuated filament, the end being blunt (H). A well developed palmate will usually have 10 or more leaflets like those just described (E). A rudimentary palmate hair (F) commonly has two to eight leaflets, lanceolate in shape, i.e., not very wide and with few or no notches.

Next in importance are the lateral hairs (Hairs No. 6) on abdominal segments III-VI; see Fig. 103. These are usually the longest hairs on the segment. Their length in relation to each other, number and mode of branching are useful characters. The antepalmate hairs (Hairs No. 2) particularly on abdominal segment VII are utilized for identification.

The spiracular apparatus on VIII (Fig. 127) consists of a central depression, the respiratory fossa, in which the spiracles are located. The spiracles constitute the only point of intake for atmospheric air required by the larva in respiration. Surrounding the fossa are four protective structures, the fan-shaped plate (ap) anteriorly, the two lateral papillae (lpa), and the posterior, concave scoop (sc). The anterior plate sometimes bears a long, thin, appendage, the stigmal club (scl), which extends posteriorly between the spiracles and overlies the scoop. The scoop has two lateral plates, forming the side walls, and a median plate, heavily sclerotized anteriorly.

On either side of the spiracular apparatus are strongly sclerotized plates, the combs, or pecten. In the Philippine anopheline larvae the posterior edge of the comb carries a number of teeth, usually several long and many more distinctly short. However, they may be subequal or have graduated lengths. Their bases ordinarily have fine serrations.



3(2).	Palmate hairs on abdominal segment I hair-like, simple or with two or three branches (Fig. 109) (<i>lindesayi</i> species group)
	Palmate hairs on abdominal segment I well developed (Fig. 110) (<i>aitkenii</i> species group)
	36 5 1 1 3 5 6° 1 1 5 6°
	Figure 109. (135x)
	3^{6} 3^{1} 3^{1} 1^{1
4(3)	Leoflete of abdominal national hairs without filaments
-(3).	(Fig. 111)
	Leaflets of abdominal palmate hairs with distinct filaments (Fig. 112)

Figure 111. (350x)

Figure 112. (350x)



7(6).



Figure 119. (220x)



acaci

8

aitkenii

bengalensis

Figure 120. (220x)

8(7).

Inner clypeal hairs with two or three branches, all beginning about one-fourth of the distance from base (Fig. 121)

Inner clypeal hairs usually with four to seven branches, if two- or three-branched, then branches beginning one-third or farther from base (Fig. 122)



Figure 121. (220x)



Figure 122. (250x)

9(2).	Palmate hairs undeveloped on thorax and abdomen, branches hair-like (Fig. 123) (umbrosus species group)	
		Palmate hairs well developed on at least some abdominal segments, branches broad, leaf-like (Fig. 124).	
		2 10 \$ 0'	2
		Figure 123. (98x)	
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		166 3 4	4 3 69
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		Figure 124. (92x)	
10(9).	Outer clypeal hairs simple or with two to five branches (Fig. 125)	samarensis
		Outer clypeal hairs dendritic, with more than 10 branches	
		(Fig. 126)	baezai
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		Figure 125. (240x)	

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24(23). Leaflets of palmate hairs on abdominal segments III-VII lanceolate; lateral hairs of abdominal segment III with fewer than 10 branches (Fig. 155) tessellatus Leaflets of palmate hairs on abdominal segments III-VII with distinct shoulders and filaments, lateral hairs of abdominal segment III with more than 10 branches (Fig. 156) . . 25 ø Ш (175x) Figure 155. E ш (160x) (110x) Figure 156. (105x) Abdominal segment II with palmate hairs moderately 25(24). developed, usually with thin lanceolate leaflets (Fig. 157) balabacensis balabacensis Abdominal segment II with palmate hairs weakly developed, branches hair-like or only slightly flattened (Fig. 158) . 26 Figure 157 Figure 158. (520x) (570x)

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^{*}Since A. sundatcus Rodenwaldt may occur in the Philippines, its larvae will come out in this key to the second part of couplet 33. It may be distinguished from vagus limosus and indefinitus by mesothoracic hair 4, which in sundaicus has three branches from near base, and in the latter two, is double, or if three-branched, the third arises from halfway along the hair.



REFERENCES:

Baisas, F.E. and F.L. Banez. 1960. Arthropods of medical importance. Inst. Malariol., Div. Malaria, Bur. Dis. Control, Dept. Hlth., Manila, Philippines, 105 pp (mimeo.).

and F.H. Dowell. 1965. Keys to the adult female and larval anopheline mosquitoes of the Philippines. Jour. Med. Ent. 4(1):11-23.

- Belkin, J.N. 1951. A revised nomenclature for the chaetotaxy of the mosquito larva. Amer. Mid. Nat. 44:678-698.
- Bohart, R.M. 1945(?) A synopsis of the Philippine mosquitoes. U.S. Naval Med. Res. Unit No. 2, Navmed 580, 88 pp.
- Borror, D.J. and D.M. Delong. 1963. An introduction to the study of insects. New York, Holt, Rinehart and Winston, 819 pp.
- Christophers, S.R. 1933. Fauna of British India, Diptera, Family Culicidae, Tribe Anophelini, Vol. IV, London, Taylor and Francis, 371 pp.
- Colless, D.H. 1956. The Anopheles leucosphyrus group. Trans. Roy. Ent. Soc. London. 108(3):37-116.
- Darsie, R.F. Jr. and A.C. Ramos. 1969. Manual of malaria entomology. 2nd Ed. Manila, Malaria Eradication Training Center. 142 pp.
- Delfinado, M.D. 1966. The culicine mosquitoes of the Philippines, Tribe Culicini (Diptera, Culicidae). Mem. Amer. Ent. Inst. No. 7, 252 pp.
 - _____, G.B. Viado and L.T. Coronel. 1962. A checklist of Philippine mosquitoes with a larval key to genera (Diptera, Culicidae). Phil. Jour. Sci. 91(4):433-457.

King, W.V. 1932. The Philippine Anopheles of the rossi-ludlowae group. Phil, Jour. Sci. 47(3):305-342.

and F.E. Baisas. 1936. A new species and a new variety of Philippine Anopheles related to Anopheles leucosphyrus Donitz. Proc. Ent. Soc. Wash. 38(5):79-89.

Mendoza, J.B. 1948. Two more Philippine Anopheles in the Myzorhynchus series. Monthly Bull. Bur. Health. 33(3):171-184.

_____ 1954a. Pictorial key to the adults (females) of Philippine Anopheles. Publ. Hlth. Res. Lab., Div. Malaria, Bur. Dis. Control, Dept. Hlth., Manila, Philippines.

_____ 1954b. Pictorial key to Philippine anopheline larvae. Publ. Hlth. Res. Lab., Div. Malaria, Bur. Dis. Control, Dept. Hlth., Manila, Philippines.

- Puti, I.M. 1960. Synoptic tables for the identification of the full-grown larvae of the Indian anopheline mosquitoes. Indian Hlth. Bull. 16, 7th Ed., 104 pp.
- Reid, J.A. 1966. A note on Anopheles subpictus Grassi and A. indefinitus Ludlow (Diptera: Culicidae) Jour. Med. Ent. 3(3-4):327-331.

Russell, P.F. and F.E. Baisas. 1934. A practical illustrated key to larvae of Philippine Anopheles. Phil. Jour. Sci. 55(4):305-336.

Jour. Sci. 59(1):15-64. Jour. Sci. 59(1):15-64.

L.S. West, R.D. Manwell and G. MacDonald. 1963. Practical malariology; 2nd Ed. London, Oxford Univ. Press, 750 pp.

Simmons, J.S. and T.H.G. Aitken. 1942. The anopheline mosquitoes of the northern half of the Western Hemisphere and of the Philippine Islands, U.S. Army Med, Bull, No. 59, 213 pp.

Snodgrass, R.E. 1959. The anatomical life of the mosquito. Smiths. Misc. Coll. 139(8):1-87.